

Standard Test Method for Nitrite-Nitrate in Water by Nitrate Reductase¹

This standard is issued under the fixed designation D7781; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ε) indicates an editorial change since the last revision or reapproval.

1. Scope

- 1.1 This test method is applicable to the determination of nitrate plus nitrite (as nitrogen) in drinking water, surface, saline, wastewater, and ground waters. The applicable range of this test method is from 0.05 to 5 mg/L of nitrogen. The range may be extended upward by dilution of an appropriate aliquot. The 40 CFR Part 136 Method Detection Limit (MDL) is 0.02 mg /L.
- 1.2 It is the user's responsibility to ensure the validity of this test method for waters of untested matrices. The quality control criteria in Section 17 for method blanks, laboratory control samples, matrix spikes and matrix duplicates must be met.
- 1.3 The values stated in SI units are regarded as standard. No other units of measurement are included in this standard.
- 1.4 This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.

2. Referenced Documents

2.1 ASTM Standards:²

D992

D1129 Terminology Relating to Water

D1141 Practice for the Preparation of Substitute Ocean Water

D1193 Specification for Reagent Water

D1254 Method of Test for Nitrite Ion in Water (Withdrawn 1980)³

D3867 Test Methods for Nitrite-Nitrate in Water

D5810 Guide for Spiking into Aqueous Samples

D6146 Guide for Monitoring Aqueous Nutrients in Watersheds

3. Terminology

- 3.1 *Definitions*—For definitions of terms used in these test methods, refer to Terminology D1129.
 - 3.2 Definitions of Terms Specific to This Standard:
- 3.2.1 *bispecific NaR*, *n*—nitrate reductase that can use either NADH or NADPH as its electron donor (cofactor).
- 3.2.2 discrete analyzer, n—a programmable, computer-controlled instrument that automates wet-chemical analysis by using one or more robotic arms interfaced to high-precision volumetric dispensers to aspirate and dispense samples, standards, diluents, and reagents.
- 3.2.3 *Greiss reaction, n*—chemical formation of an azo dye by diazotization of nitrite ion with sulfanilamide and subsequent coupling with N-(1-naphthyl)ethylenediamine hydrochloride.
- 3.2.4 *NADH*, *n*—nicotinamide adenine dinucleotide, reduced form is a coenzyme found in all living cells.
- 3.2.5 *NADPH*, *n*—nicotinamide adenine dinucleotide phosphate, reduced form is a coenzyme found in all living cells; NADP+ is the oxidizing form and NADPH is the reducing form.
- 3.2.6 *nitrate reductase (NaR)*, *n*—NADH:NaR (EC1.7.1.1 and CAS 9013-03-0) or bispecific NaR (EC 1.7.1.2 and CAS 9029-27-0) with 1 unit of enzyme activity defined as 1 micromol nitrite produced per minute at 30°C, at pH 7 with NADH (refer to 3.2.4 and 10.2) as an electron donor.

4. Summary of Test Method

- 4.1 *Nitrite-Nitrate Nitrogen*—The sample is mixed with a buffered solution containing NAD(P)H: nitrate reductase (EC 1.7.1-3) and NADH or NADPH to reduce nitrate ion to nitrite ion. The combined nitrite-nitrate (expressed as mg/L NO₃+NO₂-N) is determined by diazotizing the total nitrite ion with sulfanilamide and coupling with *N*-(1-naphthyl) ethylene-diamine dihydrochloride to form a highly colored azo dye that is measured spectrophotometrically at about 540 nm.
- 4.2 Nitrite Nitrogen—The nitrite ion (expressed as mg/L NO₂-N) originally present in the sample can be determined separately by carrying out the procedure and omitting the reduction step.

¹ This test method is under the jurisdiction of ASTM Committee D19 on Water and is the direct responsibility of Subcommittee D19.05 on Inorganic Constituents in Water.

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² For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

³ The last approved version of this historical standard is referenced on www.astm.org.

4.3 Nitrate Nitrogen—The nitrate ion as nitrogen can be calculated as the difference between the combined nitrate plus nitrite (NO_3+NO_2-N) and the nitrite (NO_2-N):

$$mg/LNO_3 - N = mg/L(NO_3 + NO_2 - N) - mg/L(NO_2 - N)$$
(1)

5. Significance and Use

5.1 This test method replaces Test Methods D1254 (Nitrite) and D992 (Nitrate). The nitrite test method (Test Method D1254) used a reagent that is considered to be a potential carcinogen. The nitrate test method (Test Method D992) has been shown to have relatively large errors when used in wastewaters and also has greater manipulative difficulties than the test method described herein.

5.2 This test method can be used in place of Test Methods D3867 (Nitrite-Nitrate). Test Methods D3867 uses cadmium for the reduction of nitrate to nitrite. Cadmium is considered a toxic metal. Also, the heterogeneous cadmium reductant creates greater difficulty than the reduction described in this test method.

6. Interferences

- 6.1 Turbid samples should be filtered prior to analysis to eliminate particulate interference.
- 6.2 Sample color that absorbs at wavelengths between 520 and 540 nm interferes with the absorbance measurements. When color is suspect, analyze a sample blank, omitting the N-(1-naphthyl)ethylenediamine dihydrochloride from the color reagent.

Note 1—The instrumentation described in this standard may automatically correct for some turbidity and sample color. See the instrument manual for further information.

6.3 Certain ions may cause interferences. See Table 1.

7. Purity of Reagents

- 7.1 Reagent grade chemicals shall be used in all tests. Unless otherwise indicated, it is intended that all reagents shall conform to the specifications of the Committee on Analytical Reagents of the American Chemical Society, when such specifications are available.⁴ Other grades may be used, provided it is first ascertained that the reagent is of sufficient high purity to permit its use without lessening the accuracy of the determination.
- 7.2 Purity of Water—Unless otherwise indicated, references to water shall be understood to mean reagent water conforming to Specification D1193, Type I or Type II. Other reagent water types may be used, provided it is first ascertained that the water is of sufficiently high purity to permit its use without adversely affecting the bias and precision of these test methods.

8. Sampling and Sample Preservation

8.1 Collect the sample in accordance with Guide D6146.

TABLE 1 Determination of Nitrate in the Presence of Potential Interferences

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Species	Concentration Added (mg/L)	Unspiked Sample Result (mg/L)	Spiked Sample Results (mg/L)	Spike Added (mg/L)	% Recovery
Cl ⁻	500	0.02 0.17	0.23 2.54	0.200 2.50	105 95
F-	500	0.01	0.22	0.200	105
Br⁻	500	<0.01 0.15	0.21 2.65	0.200 2.50	100 100
PO ₄ -3	500	0.01 0.14	0.22 2.54	0.200 2.50	105 96
SO ₄ -2	500	<0.01 0.14	0.21 2.53	0.200 2.50	105 96
Fe	500 1.0	0.17 <0.01 0.168	2.60 0.21 2.59	2.50 0.200 2.50	97 105 96
Zn	1.0	<0.01 0.14	0.22 2.64	0.200 2.50	110 100
Al	1.0	<0.01 0.14	0.21 2.53	0.200 2.50	105 96
BrO ₃ -	1.0	<0.01 0.17	0.22 2.64	0.200 2.50	110 99
CIO ₂ -	1.0	0.01 0.14	0.22 2.54	0.200 2.50	110 96
CLo ₃ -	1.0	0.23	2.45	2.50	89
CHCL ₃	>Miscibility	<0.01	0.21	0.200	105

8.2 When nitrite ion is to be determined separately, analyze within 48 hours after sampling. Even when sterile bottles are used, bacteria naturally present in the water may cause conversion of all or part of nitrite ion to other forms such as nitrate or ammonia. Ammonia and natural amines, which are frequently present in natural waters, may react with nitrites to form nitrogen. If samples are to be stored for 48 h or less, preserve the sample by refrigeration at 2–6°C. If the sample must be stored for more than 48 h, preserve it by the addition sulfuric acid to pH 2 in addition to refrigeration at 2–6°C.

Note 2—Use sulfuric acid for preservation of nitrite-nitrate nitrogen only. Samples for nitrite must be analyzed within 48 hours.

Note 3—Sulfuric acid does not necessarily inhibit oxidation and mercury compounds should be avoided to prevent environmental pollution

Note 4—Residual chlorine does not interfere, however, attempts to remove residual chlorine (such as addition of ascorbic acid) interfere by inhibiting reduction of nitrate to nitrite. Do attempt to remove residual chlorine.

9. Apparatus

9.1 Automated discrete analysis system (see 3.2.2).

10. Reagents

10.1 *Phosphate Buffer Solution*—Dissolve 3.75 g of potassium dihydrogen phosphate (KH2PO4), 0.01 g of disodium ethylenediaminetetraacetate dehydrate

⁴ Reagent Chemicals, American Chemical Society Specifications, American Chemical Society, Washington, DC, www.chemistry.org. For suggestions on the testing of reagents not listed by the American Chemical Society, see the *United States Pharmacopeia and National Formulary*, U.S. Pharmacopeial Convention, Inc. (USPC), Rockville, MD, http://www.usp.org.

TABLE 2 Example Concentrations of Calibration Standards

NO ₃ -N or NO ₂ -N, mg/L	mL of 10 mg/L Standard Solution/100 mL final volume
0.05	0.5
0.1	1.0
0.5	5.0
1.0	10
2.0	20
3.0	30
5.0	50

TABLE 3 Reduction Efficiency

NO ₃ -N or NO ₂ -N, mg/L	mL of 10 mg/L Standard Solution/100 mL final volume		
Mean (%)	103		
Standard Deviation	4.14		
Lower Limit (%)	91		
Upper Limit (%)	115		

 $(C_{10}H_{14}O_8N_2Na_2\cdot 2H_2O)$, and 1.4 g potassium hydroxide (KOH) in about 500 mL reagent water contained in a 1000 mL volumetric flask, dilute to the mark and mix. Transfer this solution to a screw-cap container and store at 2–6°C. This solution is stable for 6 months.

10.2 β-nicotinamide adenine dinucleotide, Reduced Form (NADH) Stock Solution (2 mg/mL)—Dissolve 0.1 g NADH ($C_{21}H_{27}N_7O_{14}P_2$) in 25 mL of reagent water contained in a 50 mL volumetric flask, dilute to the mark and mix. Transfer 1-mL aliquots to 1.5 mL snap-cap colorless polypropylene vials and store at $-20^{\circ}C$. Stable for 1 month.

Note 5—NADH is a hygroscopic white powder that is freely soluble in water. The solids are stable if stored dry and protected from light. Neutral solutions are colorless and stable for 1 week if stored at 4°C, but decompose rapidly under basic or acidic conditions.

10.3 NADH Working Solution—Thaw one 1-mL vial of NADH stock (refer to 10.2) and dilute to 10 mL with phosphate buffer (refer to 10.1). This reagent is stable for about 8 hours. Prepare sufficient NADH working solution for the number of samples and standards to be analyzed.

Note 6—NADH inhibits color formation in the Greiss reaction (refer to 3.2.3). The molar concentration of NADH in the reduction medium should be about twice that of the highest calibration standard.

10.4 Sulfanilamide (SAN) Reagent (10g/L)—While stirring constantly add 300 mL of concentrated hydrochloric acid (HCl, 37 % w/v)) and 10 g of sulfanilamide (SAN, C₆H₈N₂O₂S) to about 500 mL reagent water contained in a 1000 mL volumetric flask, dilute to the mark and mix. This solution is stable for about six months when stored in a brown bottle at 20°C.

10.5 *N-(1-naphthyl)ethylenediamine* dihydrochloride (*NED*) Solution (1g/L)—dissolve 1 g NED (C₁₀H₇NHCH₂CH₂NH₂·2HCl) in about 500 mL reagent water contained in a 1000 mL volumetric flask, dilute to the mark and mix. Transfer to a glass or amber screw-cap container. This solution is stable for 6 months at 20°C.

10.6 *Nitrate Reductase (NaR)*—Follow the manufacturer's instructions for preparing a solution of 1 unit NaR (refer to 3.2.6) activity per mL of phosphate buffer (refer to section 10.1). Dilute 3 units NaR to 20 mL with phosphate buffer. Store

the solution at 2–6°C, where it is stable for 8 hours. Prepare sufficient NaR for the total number of samples and standards to be analyzed.

Note 7—For some NaR forms, high phenolic content humic substances (>2 mg dissolved organic carbon /L) have little affect on the NaR activity in the temperature range of 5–15°C, but become increasingly inhibitory in the temperature range of 20–30°C. Humic substances at the operation temperatures specified in this standard do not inhibit other forms of NaR.⁵ If humic acids are expected to be present the user must verify reduction efficiency of the NaR is use by analysis of Quality Control checks that approximate the sample matrix.

10.7 Nitrate Solution, Stock (1000 mg/L NO₃-N)—Dry potassium nitrate (KNO₃) in an oven at 105°C for 24 h. Dissolve 7.218 g in water in about 500 mL reagent water contained in a 1000 mL volumetric flask, dilute to the mark and mix. This solution is stable for up to 2 months with refrigeration. Alternatively, certified nitrate stock solutions are commercially available through chemical supply vendors and may be used.

10.8 Nitrate Solution, Standard (10 mg/L NO_3 -N)—Dilute 10 mL of stock nitrate solution (10.7) to 1 L with water and store in a dark bottle. Prepare fresh as needed.

10.9 Nitrite Solution, Stock (1000 mg/L NO₂-N)—Place about 7 g of potassium nitrite (KNO₂) in a tared 125-mL beaker and dry for about 24 h to a constant weight in a desiccator containing a suitable desiccant. Adjust the weight of the dry potassium nitrite to 6.072 g. Add 50 mL of water to the beaker, stir until dissolved, and transfer quantitatively to a 1000-mL volumetric flask. Dilute to the mark with water store in a sterilized bottle under refrigeration. Prepare fresh as needed. Alternatively, certified nitrite stock solutions are commercially available through chemical supply vendors and may be used.

Note 8—Potassium nitrite is easily oxidized; use only dry, free flowing white, or yellowish white crystalline powder of this reagent.

10.10 Nitrite Solution, Standard (10 mg/L NO₂-N)—Dilute 10 mL of stock nitrite solution (10.9) to 1 L with water. This solution is unstable; prepare fresh as needed.

11. Hazards

11.1 All reagents and standards should be prepared in volumes consistent with laboratory use to minimize the generation of waste.

12. Calibration

12.1 Using the standard nitrate solution (10.8) prepare calibration standards by using the automated calibration function of the discrete analyzer (3.2.2). Table 2 specifies suggested calibration levels.

12.2 Prepare at least one calibration standard from the standard nitrite solution (10.10) at the same concentration as one of the nitrate standards to verify the efficiency of the reduction. Verify that reduction efficiency is between 90 and 115 % with each batch of enzyme. See Table 3.

Note 9—When the sample to be analyzed is saline water, use substitute ocean water (SOW) to prepare the standards (Practice D1141 or a

⁵ NaR available from the Nitrate Elimination Company Inc. (NECi), www.nitrate.com, has been found suitable.